Introduction

Clonal hematopoiesis, as determined by the presence of TET2, ASXL1, and DNMT3 mutation, is believed to be increasingly common in normal individuals as they age. This phenomenon is currently referred to as clonal hematopoiesis of indeterminate potential (CHIP). It has been reported that a few of these patients will develop myeloid neoplasms that begin as myelodysplastic syndrome (MDS) and may evolve into acute myeloid leukemia (AML). However, the clinical relevance of this particular subset of gene mutations (TET2, ASXL1, and DNMT3A) in healthy individuals is unclear as there are more than 50 genes that have been reported to be involved in myeloid neoplasms. FLT3 and NPM1 mutations have been reported in de novo AML patients and may occur as a secondary mutations on an MDS background. In order to clarify the significance of TET2, ASXL1, and DNMT3 in CHIP and their relationship to MDS and AML, we assessed the prevalence of TET2, ASXL1, and DNMT3 genes in patients with NPM1 and FLT3 mutations. In addition, since IDH1/2 gene are rarely mutated with TET2, we also studied the frequency of mutations of various myeloid genes in this group of patients.

Methods

A total of 6390 consecutive bone marrow aspirate samples or peripheral blood samples submitted with clinical impression of AML, MDS, or MPN between January 2014 and mid 2017 were tested for mutations in myeloid genes using next generation sequencing (NGS). We used the TruSight Myeloid Panel (Illumina, San Diego, CA) for detecting missense mutations and fragment length analysis (FLA) for detecting ITD in FLT3 and large indels in CALR. DNA was extracted from samples using the QIAamp DNA Mini Kit. This NGS testing covers mutations in 54 myeloid-related genes. The average depth of sequencing was 10,000x.

Results

In our database of consecutively tested patients, there were 311 patients with *FLT3* mutations, 318 patients with NPM1 mutation, and 467 patients with IDH1/2 mutation. In addition, there were 308 patients diagnosed as MDS. All these patients tested for mutations in all 54 myeloid-related genes. The median age of patients in FLT3, MDS, IDH1/2, and NPM1 groups was 65, 75, 69, and 66, respectively. The top 4 most frequently mutated in genes in the FLT3-positive patients, which present AML, were NPM1, DNMT3, WT1, and RUNX1. In patients with NPM1 mutations, also representing AML, the top 4 most mutated genes were FLT3, IDH2, NRAS, and PTPN11. In the IDH1/2 mutated patients, the top 4 mutated genes were DNMT3A, SRSF2, ASXL1, and NPM1. As expected in the MDS group, the top 4 mutated genes were TET2, DNMT3A, ASXL1, and SRSF2.

Clonal Hematopoiesis in Normal Individuals is Not Random and Likely Reflects Early MDS

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Frequency of Co-mutated Genes in Sub-Cohorts: NPM1+, FLT3+, IDH1/2+, and MDS

Mutateo Gene	CHIP: VAF<20% VAF	CHIP: VAF<20% VAF	
DNMT3/	A <i>32.88%</i>	51.87%	
TP53	22.71%	35.83%	
SF3B1	18.31%	28.88%	
TET2	16.27%	25.67%	
ASXL1	14.92%	23.53%	
JAK2	5.93%	9.36%	
U2AF1	4.92%	7.75%	
MYD88	4.24%	6.68%	
CEBPA	2.71%	4.28%	
KRAS	2.71%	4.28%	
RUNX1	2.54%	4.01%	
BRAF	2.20%	3.48%	
IDH2	2.20%	3.48%	
NRAS	2.20%	3.48%	
FLT-3	2.03%	3.21%	
BCOR	1.53%	2.41%	
SRSF2	1.53%	3.21%	
ZRSR2	1.36%	2.14%	
IDH1	1.19%	1.87%	

NPM1-Positive		FLT-3-Positive		IDH1/2-Positive		MDS	
N=318		N=311		N=467		N=308	
Gene	%	Gene	%	Gene	%	Gene	%
DNMT3A	47.81%	NPM1	44.09%	DNMT3A	34.26%	TET2	31.29%
FLT3-ITD	30.31%	DNMT3A	40.26%	SRSF2	27.62%	DNMT3A	28.39%
FLT3-KD	20.31%	TET2	17.25%	ASXL1	22.48%	ASXL1	25.16%
IDH2	19.06%	WT1	14.70%	NPM1	22.06%	SRSF2	17.10%
NRAS	17.81%	RUNX1	14.38%	RUNX1	19.70%	SF3B1	14.52%
PTPN11	16.56%	ASXL1	12.46%	NRAS	9.64%	RUNX1	10.97%
IDH1	15.00%	IDH2	11.18%	TET2	7.71%	TP53	10.32%
TET2	14.38%	NRAS	11.18%	FLT3-ITD	7.28%	NRAS	7.74%
WT1	6.56%	IDH1	6.71%	BCOR	6.64%	U2AF1	5.81%
KRAS	5.94%	PTPN11	6.39%	JAK2	6.64%	IDH2	5.48%
SRSF2	5.31%	BCOR	5.75%	TP53	6.64%	JAK2	5.48%
ASXL1	5.00%	CEBPA	4.47%	STAG2	6.21%	SETBP1	4.84%
CEBPA	4.69%	STAG2	3.83%	PTPN11	5.35%	ZRSR2	4.84%
SF3B1	2.50%	SF3B1	3.51%	FLT3 KD	4.93%	CBL	4.19%

op table shows the frequency of nutations in various genes detected patients with CHIP. The bottom ables show the frequency of conutated genes in various groups. The NPM1+ (318) and FLT3+ (311) epresent AML. The IDH1/2+ (467) epresenting confirmed myeloid neoplasms. The MDS (308) group vho were confirmed by VAF>20%. As hown, the most commonly mutated genes in CHIP as the same genes nutated in MDS. In contrast, patients vith *FLT-3* and *NPM1*, which are more ikely to represent AML as well as patients with IDH2/1 are more likely to nave mutation pattern different from hat of CHIP and MDS.

Variation in Age Between Various Groups

Groups	Valid N	Mean	Median
FLT3	311	62	65
MDS	308	72	75
Normal	332	63	65
IDH	467	68	69
NPM1	318	64	66
CHIP	590	67	70

Key Points

- is <20%.
- Patients samples were referred for molecular testing on patients with clinical suspicion of hematologic neoplasm due to anemia, thrombocytopenia, or neutropenia.
- A total of 7497 samples were tested between 2014 and mid 2017. Of these 4075 had one mutation in one or more genes.
- Only 590 of 7497 (8%) total tested and 14% of cases with mutations showed a mutation in one gene with VAF<20%. Of these 374 (63%) had VAF <10%.
- Mutation pattern in cases with CHIP in this population of patients, who have one or more cytopenia, is more suggestive of early MDS.
- Unusually high rate of TP53 mutation is detected in this group of patients.
- A high percentage of patients have MYD88 mutations, suggesting lymphoid rather than myeloid neoplasm in this sub-cohort.

Conclusion

The demonstration that mutations in TET2, DNMT3A, and ASXL1 are most common in patients presenting with MDS, as well as in CHIP, but not in patients presenting with acute myeloid neoplasms, suggests that CHIP likely represents early MDS. It is possible that mutations in these genes in few hematopoietic cells may not be adequate for manifestation of clinical MDS, but the presence of these mutations in large number of cells (high VAF) or the accumulation of mutations in additional genes is necessary for clinical disease. Therefore the clinical relevance of CHIP should always be considered in conjunction with other mutations and with VAF.

• A few of patients with CHIP will develop myeloid or lymphoid neoplasms.

• We considered a patient having CHIP when a mutation in one gene is detected and the VAF