

Poster Number: 8517 Presenting author: davina.gale@cruk.cam.ac.uk

Davina Gale^{1,2}, Katrin Heider^{1,2}, Malcolm Perry³, Sophie Hackinger³, Giovanni Marsico³, Andrea Ruiz-Valdepenas^{1,2}, Viona Rundell⁴, Jerome Wulff⁴, Garima Sharma³, Karen Howarth³, David Gilligan^{5,6}, Susan V. Harden^{5,*}, Doris M. Rassl^{6,2}, Robert C. Rintoul^{7,6,2}, Nitzan Rosenfeld^{1,2,3}

Cancer Research UK Cambridge Institute, University of Cambridge, Robinson Way, Cambridge CB2 ORE; ³Inivata Ltd, The Glenn Berge Building, Babraham Research UK Cambridge, UK; ⁴Cambridge UK; ⁴Cambridge CB2 ORE; ³Inivata Ltd, The Glenn Berge Building, Babraham Research UK Cambridge, UK; ⁴Cambridge, UK; ⁴Cambridge CB2 ORE; ³Inivata Ltd, The Glenn Berge Building, Babraham Research UK Cambridge, UK; ⁴Cambridge CB2 ORE; ⁴Cambridge, UK; ⁴Cambridg Clinical Trials Unit – Cancer Theme, Cambridge, Addenbrooke's Hospital, Cambridge CB2 0QQ; ⁵Addenbrooke's Hospital, Cambridge CB2 0QQ; ⁵Addenbrooke's Hospital, Cambridge CB2 0AY; ⁷Department of Oncology, University of Cambridge Hutchison–MRC Research Centre, Box 197, Cambridge, Biomedical Campus, Cambridge, CB2 0XZ, UK Current affiliation: Cancer Research Programme, School of Public Health and Preventive Medicine, Monash University, Melbourne 3004, Australia

INTRODUCTION

- Identification of minimal residual disease (MRD) in patients with localized non-small cell lung cancer (NSCLC) following treatment with curative intent holds promise for identifying patients who are at higher risk of relapse who may benefit from adjuvant therapy.
- Liquid biopsies based on circulating tumor DNA (ctDNA) analysis are being investigated for detection of residual disease and recurrence.
- Due to low ctDNA levels in early-stage disease or post-treatment, effective methods require high analytical sensitivity to detect variant allele fractions (VAF) below 0.01%.
- Here, we have evaluated detection of ctDNA in serial plasma samples collected from the LUCID (LUng cancer - Circulating tumor DNA) study using a personalized sequencing assay.

STUDY OBJECTIVE

The objective of this study was to test the feasibility and prognostic value of detecting ctDNA at or before relapse in 88 stage IA - IIIB NSCLC patients following treatment with curative intent, either surgery (n=69) or radiotherapy (RT) ± chemotherapy (n=19) [Table 1].

		Characteristics
Characteristics	Patients (n=88)	Histology
Age, Median (range)		Adenocarcinoma
Stage I	73 (52-88)	Squamous cell carcinoma
Stage II	74 (57-83)	Other
Stage III	67 (44-78)	Pathology
Sex		
Male	45 (51.1%)	Stage I
Female	43 (48.9%)	Stage II
Smoking status		Stage III
Never	8 (9,1%)	Treatment
Fx-smoker	63 (71 6%)	Surgery
Smoker	16 (18 2%)	ChemoRadiation
Concer bistory	20 (22%)	Time points
Lancer history	29 (33%)	Baseline
		Follow-up

Table 1: Patient demographics of NSCLC patients enrolled in the study who had available tumor tissue for exome sequencing & an available ctDNA assay (n=88).

- Plasma samples (n=363) were collected before and after treatment, and at 3, 6 and 9 months. For 17 patients, additional plasma was collected at disease relapse. Patients were followed for a median of 3 years (range: 9 months to 5 years) and disease outcomes recorded.
- Plasma ctDNA was analysed using RaDaRTM assays that target up to 48 tumor-identified variants per patient. Detection of residual ctDNA after end of treatment was compared to outcomes including Relapse Free Survival (RFS) and Overall Survival (OS).

tumor tissue & biopsy **DNA extraction**

to confirm somatic mutations and exclude clonal hematopoiesis (CHIP)



0.0007% to >2%)



Residual ctDNA after treatment predicts early relapse in patients with early-stage NSCLC



Plasma samples were analysed and ctDNA detection compared to outcomes. ctDNA Ê negative Library preparation zh-depth seauencir Assess prognostic value of iant confirmation 8 ctDNA detection according to CHIP filtering clinical outcomes

ctDNA detection prior to treatment

ctDNA was detected prior to treatment in 87%, 77% and 24% of patients with stage III, II and I disease respectively. Overall, ctDNA was detected in 26% of all samples collected at baseline and follow-up at a median variant allele fraction (VAF) of 0.047% (range:



Detected

Figure 3: ctDNA levels (VAFs), measured in parts per million (ppm), detected in plasma samples prior to treatment.



Dotted lines indicate variant allele



fractions of 0.01% and 0.1%.

Longitudinal monitoring for residual disease and recurrence

Figure 4: Examples of longitudinal monitoring of ctDNA

- Patient 1: ctDNA detected before surgery, but not at 20 days after surgery, and detected at day 174, 222 days prior to clinical progression.
- Patient 2: ctDNA detected at all timepoints, lead time 203 days
- Patient 3: ctDNA detected before and after surgery, but not after adjuvant chemotherapy. No progression in 3 years of further follow-up.
- Patient 4: ctDNA detected before and after surgery, and at all follow-up time points, many months prior to clinical progression.



Figure 5: Heatmap showing analysis of tumor, buffy coat and plasma samples from Patient 4 using a patient-specific assay. Each row represents a different sample type and each column is a different variant. Variants shaded in grey were excluded from analysis due to absence in tumor DNA or presence in buffy coat DNA

CONCLUSIONS

cfDNA 101 days -

2012 2012

cfDNA 192 day

Our results demonstrate the ability to monitor ctDNA in NSCLC patients at or prior to relapse using a sensitive patient-specific plasma sequencing assay. ctDNA was detected in plasma samples at levels as low as 7 ppm. ctDNA detection within the landmark timepoint (2 weeks to 4 months after treatment end), was associated with shorter Relapse Free Survival (HR 14.8, p-value <10⁻⁵) and Overall Survival (HR 5.48, p-value<0.0003). In patients who progressed, detection of ctDNA preceded clinical progression by a median lead time of 212.5 days. Our results support emerging evidence that ctDNA monitoring can reliably detect residual disease in patients treated with curative intent, many months before clinical progression, and offers an opportunity to identify ctDNA-positive patients who may benefit from adjuvant therapy.



NHS **Royal Papworth Hospital**

Cambridge University Hospitals

Figure 6: Longitudinal monitoring of plasma from patients with (A) ctDNA detected (n=40) and (B) ctDNA not detected prior to treatment (n=38).





cancer (genomic investigation is underway), and one patient died of other causes. 5 patients had no samples

Figure 7: In survival analysis of 59 patients, plasma was available within a landmark timepoint, between 2 weeks and 4 months after end of initial treatment. ctDNA detection at the landmark timepoint was strongly predictive of clinical disease relapse Kaplan-Meier curve showing (A) Relapse Free Survival, Hazard Ratio: 14.8, p-value<10⁻⁵. (B) Overall Survival, Hazard ratio 5.48 (p-value<0.0003). All 10 patients with ctDNA detected at landmark had clinical progression within the study period.