

# Actionable Fusions Detected by RNA-seq Co-occur with PD-L1 Expression and Driver Mutations in Solid Tumors Patients

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## Background

Incorporating gene fusions into a comprehensive profile is critical not only because very effective therapies targeting oncogenic fusion proteins exist but they may also negate the response to therapy of an actionable SNV and InDel mutations. We examined the co-occurrence of actionable gene fusions detected by RNA-seq with actionable SNVs/InDels and with the immunotherapy response biomarker, PD-L1.

## Methods

In 2021, 5341 FFPE samples were analyzed by our clinical laboratory using a novel, clinical grade hybridization-based RNA sequencing assay that detects the expressed chimeric transcripts resulting from gene fusions (Figs. 1, 2) with 95.9% sensitivity and 100% specificity. DNA mutations (SNV/Indels) were detected with a clinical grade NGS assay and PD-L1 protein expression was determined by IHC using an appropriate LDT or FDA approved assays (SP42 or 22C3 antibodies). De-identified data were analyzed following an approved IRB protocol.

## Results

Of the 5341 patients tested for gene fusions only 0.7% were profiled with a comprehensive fusion detection panel for 250 clinically relevant fusion genes. Conversely, 67% of all patients were tested only for NTRK gene fusions. The prevalence of the following most relevant fusions was: ALK=2.31% (in particular EML4-ALK), NTRKs=0.85%, RET=0.92% and ROS1=0.80% all mostly detected in lung cancers and FGFR2=5.60% in cholangiocarcinoma and PAX-FOXO in rhabdomyosarcomas. Among NTRK fusions, NTRK3 was detected in 52.9% of the positive cases; in particular, dominant fusions are ETV6-NTRK3 (26%) and EML4-NTRK3 (20%). In the 163 fusion positive cases, 37 cases had available DNA mutation testing and PD-L1 expression testing. These patients presented 24 different actionable fusions, including ALK (2), FGFR1/2 (5), NTRK 1/2/3 (10), NRG1 (3), RET (3) and ROS1 (3) fusions. Interestingly, while 73% (27/37) of those tumors were PD-L1 positive, similar to the 75% found on the fusion negative samples, PD-L1 was positive in 88% (15/17) of the lung samples with pathogenic mutations from this subset. This was strikingly higher than the 45% (192/424) found in the fusion negative cohort. Finally, excluding TP53 mutations, NTRK fusions frequently co-occurred with RNF43, FBXW7, TERT promoter, and ARID1A pathogenic mutations. FGFR fusions co-occurred with BAP1, PBRM1 or KRAS mutations, which correlates with the fact that both FGFR2 fusions and swi/snf alterations are enriched in intrahepatic cholangiocarcinoma, where the FGFR fusions were detected. RET fusions co-occurred with ARID1A and TERT. ROS1 fusions co-occurred with SMARCA4 and KRAS pathogenic mutations simultaneously. NRG1 fusions co-occurred with ARID1A and FBXW7 mutations. EML4-ALK fusions co-occurred with FGFR2 and KMT2D variants of unknown significance.

## Conclusion

Actionable cancer driving gene fusions were detected by RNA-sequencing and co-occur with other biomarkers that can guide selection of therapies, such as immune checkpoint inhibitors (ICI) and olaparib. The data suggests that gene fusion testing is an important addition on the genomic profiling for therapy selection in solid tumors.

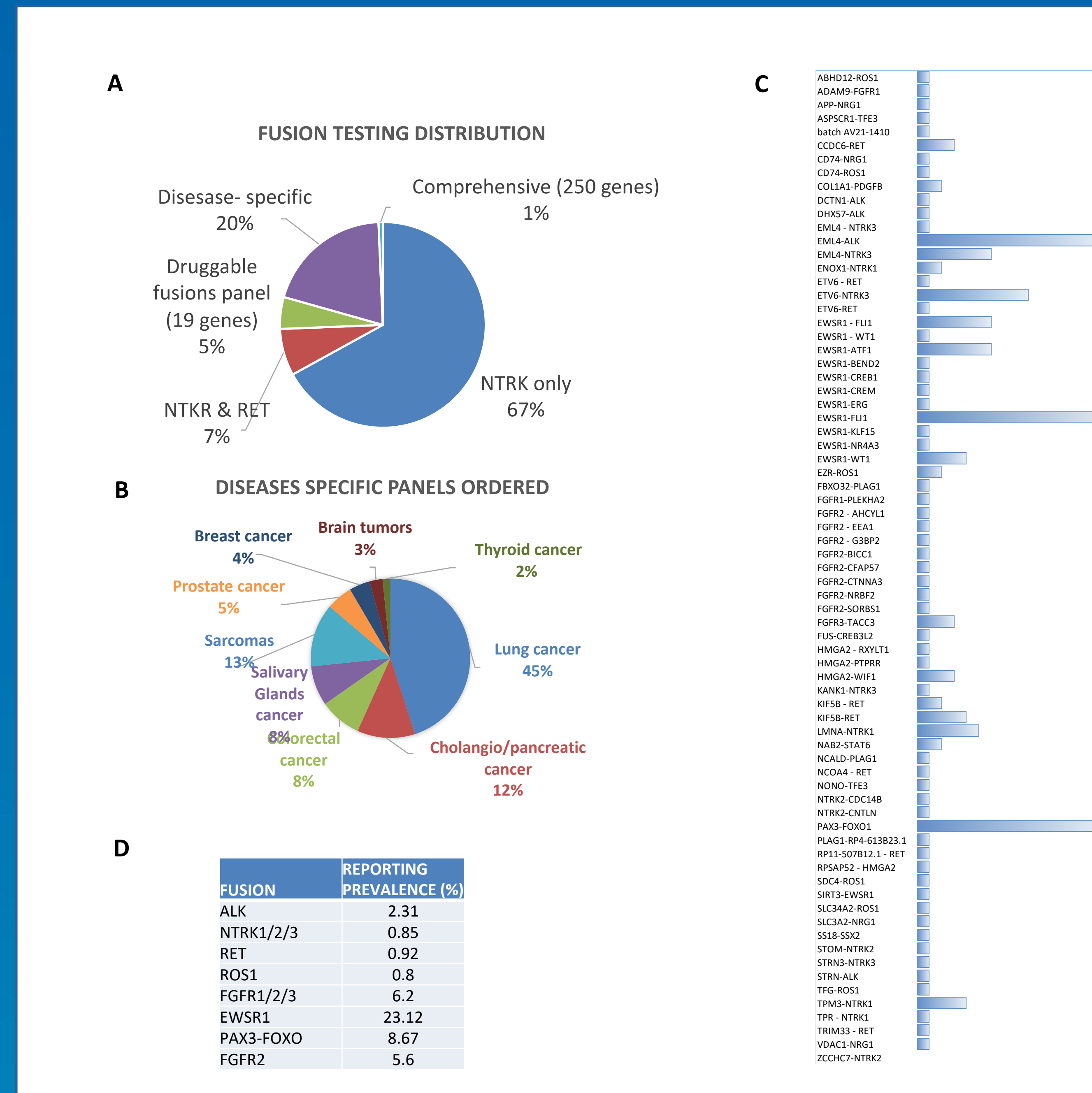
## Highlights

- Gene fusions testing in clinical practice was mainly ordered for select fusion genes
- Therapeutically actionable fusions often co-occur with other driver mutations
- Therapeutically actionable fusions are co-detected with immunosuppressive phenotype
- Gene fusions and driving mutations might jointly drive tumors biology

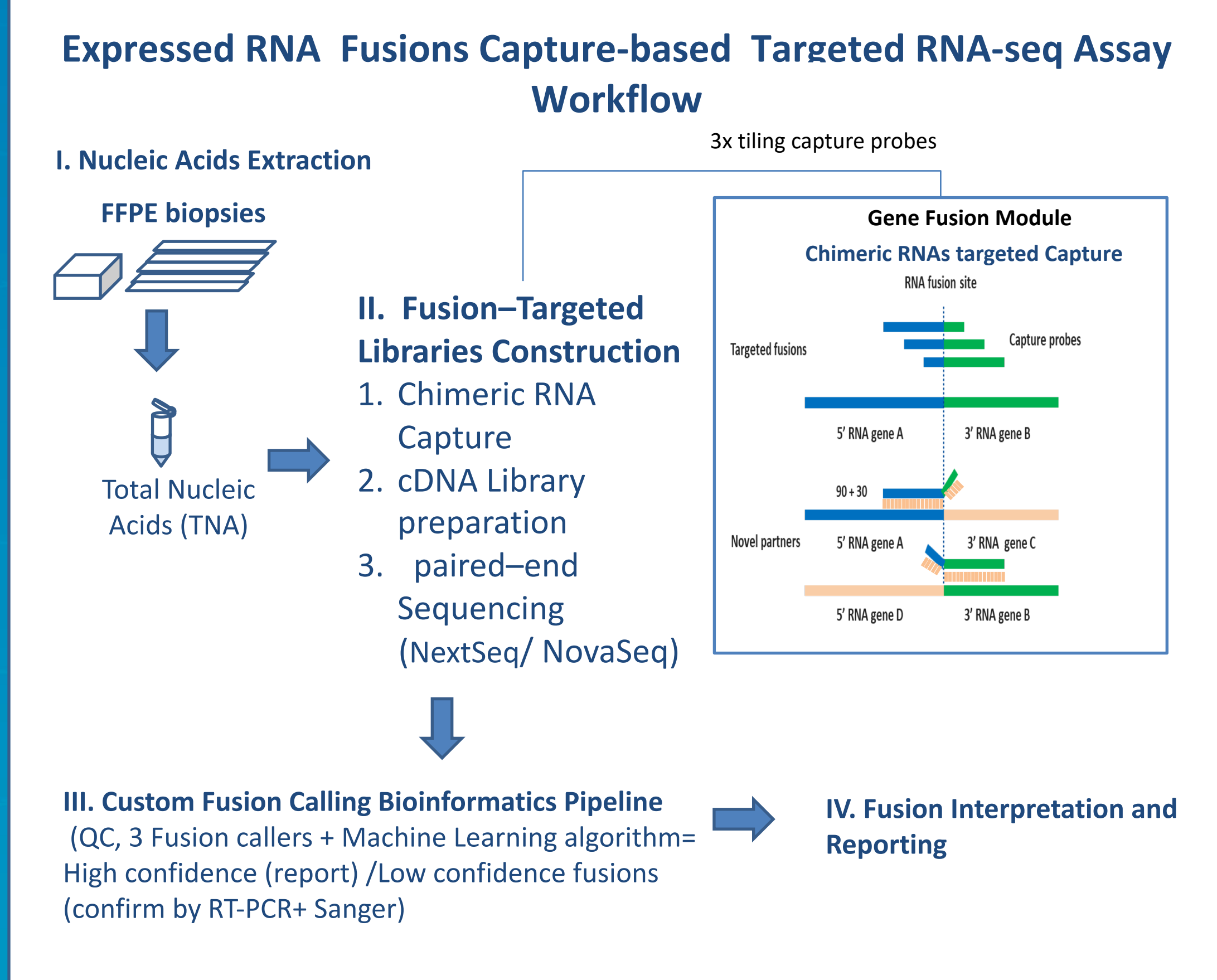
### Solid Tumor Relevant Gene Fusions and Aberrant Transcripts

250 clinically relevant fusion genes											
ABL1	C11orf95	CREB1	ESRP1	HERPUD1	MAST2	NOTCH2	PML	SET	TFE3		
ACSL3	CAMTA1	CREB3L1	ETV1	HEY1	MEAF6	NPM1	POU5F1	SLC34A2	TFG		
ACTB	CANT1	CREB3L2	ETV4	HSP1	MET	NRXN3	PRKRG	SLC34A3	THADA		
ACTL6A	CAPZA2	CREBBP	ETV5	HMG2	MET 014 *	NRG1	PRCC	SND1	THRAP3		
AFO1	CARS1	CRTC1	ETV6	HMG2046	MKRN1	NTRK1	PRKACA	SNURF	TMPP52		
AFF1	CBAT3	CRTC3	EWI1	HNRNPA2B1	MLLT1	NTRK2	PRKAR1A	SIF	TRG3		
AFF3	CCDC170	CSF1	EZR	IL2RB	MLL110	NTRK3	PRK01	SRGAP3	TPA3		
AFF4	CCDC6	CTLA4	FAM131B	IRF4	MLL11	NUP214	PRK02	SS18	TPK4		
AHHR	CCNB3	CTNNE1	FEV	ITK	MLL2	NUP98	PRK03	SSX1	TPK		
AKAP9	CEND1	CTNBL1	FGFR1	JAK2	MN1	NUTM1	PTPRK	SSX2	TRIM24		
AKT3	CND2	DOT1B	FGFR2	JAZ1	MPHSP	NUTM2A	RA251B	SSX4B	UBTF		
ALK	CND3	DOK3K	FGFR3	KAT5A	MRTF8	NUTM2B	RAF1	STAT6	USP6		
AR	CD274	DDX5	FGFR4	KDM5A	MSH2	DMD	RANBP2	STIL	VTG1A		
ARNT*	CD74	DH1	FUS	KIAA1549	MYB	PAN3	RAK1	STRN	WDR72		
ARV9*	CDH11	DNAB1	FLT3	KIF5B	MYBL1	PATZ1	BAK5E1A	SUZ12	WIF1		
ARID1A	CDK4	DUK4	FOXO1	KIT	MYC	PAX3	RET	SYK	WT1		
ASPSK1	CHD6	EGFR	FOXP1	KUJ2	MFR9	PAX7	RHEBL1	TAC1C3	WVTR1		
AT13	COR102	EGFR-RET*	FRK	KMT2A	MYLK	PAX8	ROSL	FAF15	YAP1		
ATIC	CHCHD7	ELK4	FUS	KNL1	NAB2	PBX1	RPS6K1	TAL1	YWHAE		
AXL	CIC	ELL	GUS1	KRAS	NCOA4	PCOL1	RPS3	TBL1XR1	ZNF444		
BCOR	CIT4	EML4	GLI3	LIFR	NCOA2	PDGFR	RUNX1	TEF12			
BCR	CLTC	EPC1	GLI2	LMNA	NCOA4	PDGFR	RUNX1T1	TEF3			
BRAF	CNBP	EPK5	GLI3	LIF	NRG1	PDGFR	SOX4	TEF12			
BRCA1	COA5	ERBB2	GNAS	MAML1	NFATC2	PHF1	SEC31A	TEAD1			
BRCA2	COL1A1	ERG	GOPC	MAML2	NFIB	PKCXA	SEPTINE	TEAD2			
BRD4	COL1A2	ESR1	HAS2	MAST1	NOTCH1	PLAG1	SEPTIN9	TEAD3			

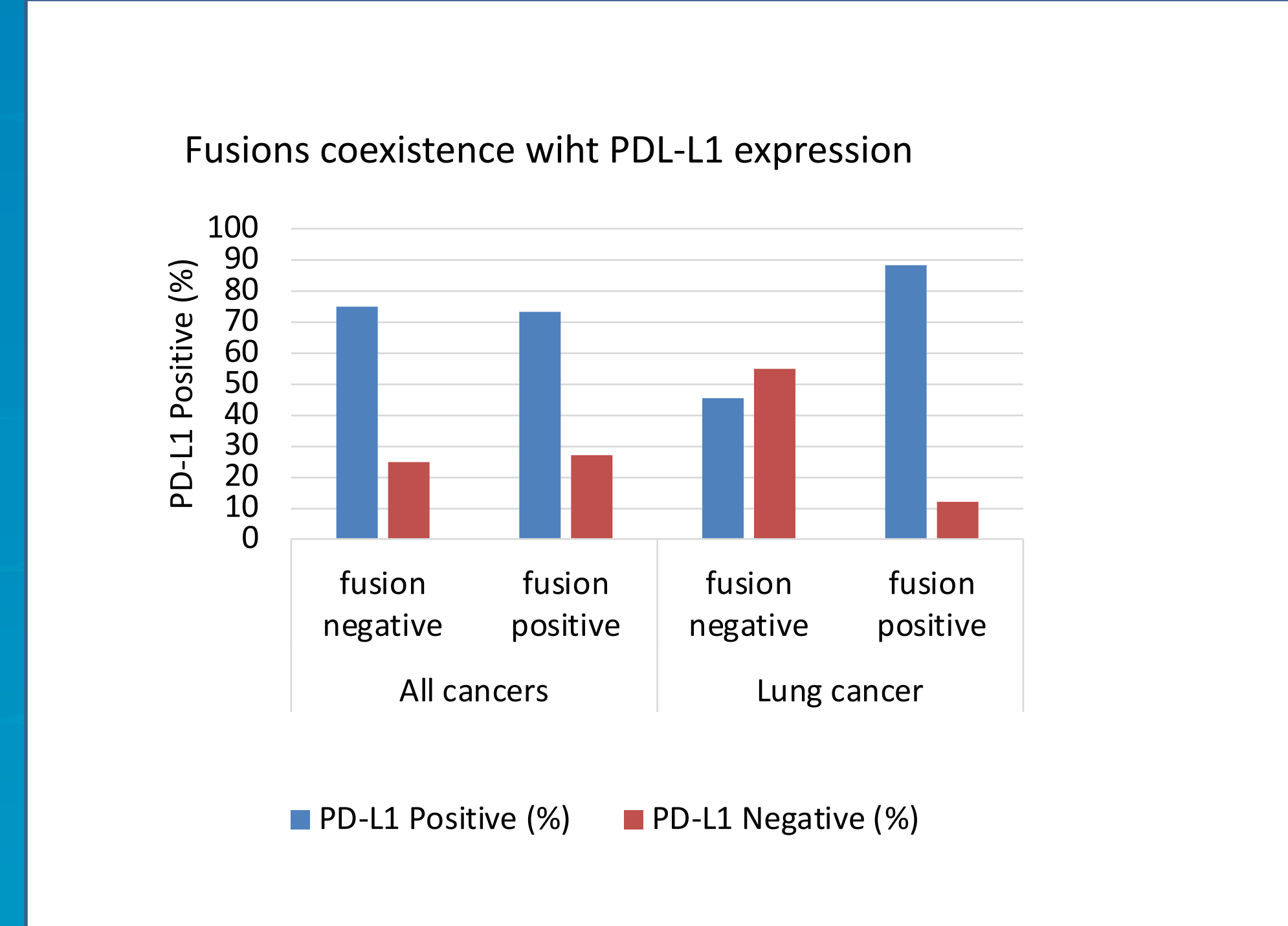
1. The fusion panel targets fusions involving 250 clinically relevant fusion genes relevant to most frequent cancers, including breast, colorectal, lung, lymphoma, pancreatic, prostate, salivary gland, sarcomas, and thyroid cancers. Fusion genes are included on NCCN and WHO guidelines, published clinical studies, and the 120 most frequent curated fusions in solid tumors from COSMIC (v91) providing >90% prevalence per fusion pair.



3. Physicians ordered fusions tests for 5341 cases. (A) All tested cases. (B) Distribution of cases tested for disease specific fusions. (C) Fusions from 163 positive reported cases. (D) Prevalence of select common fusions reported across all cancers tested.



2: A new Solid Tumor Gene Fusion RNA-Seq assay workflow. Starting with FFPE total nucleic acid extraction, the assay detects only expressed fusions through RNA fusion sequence-targeted, hybridization capture-based, stranded pair-ended RNA-sequencing on Novaseq instruments. Key step resides on custom based design of capture baits mapping to the chimeric RNA sequence as deduced from chromosomal breakpoints in annotated private and public databases.



5. Co-existence of oncogenic fusions and Immunotherapy biomarker PD-L1.

case	Fusion detected (NGS)	PD-L1 (IHC) Test Result
1	STOM-NTRK2	EXPRESSED
2	MET Exon 14 Deletion	EXPRESSED
3	KIF5B-RET	EXPRESSED
4	SLC3A2-NRG1	EXPRESSED
5	NTRK2-CDC14B	EXPRESSED
6	EML4-ALK	EXPRESSED
7	SDCA-ROS1	HIGH EXPRESSION
8	TPM3-NTRK1	HIGH EXPRESSION
9	FGFR2-AHLY1	HIGH EXPRESSION
10	KIF5B-RET	HIGH EXPRESSION
11	MET Exon 14 Skipping	HIGH EXPRESSION
12	CCDC6-RET	HIGH EXPRESSION
13	EML4-NTRK3	HIGH EXPRESSION
14	EML4-NTRK3	HIGH EXPRESSION
15	MET Exon 14 Deletion	HIGH EXPRESSION
16	EML4-NTRK3	HIGH EXPRESSION
17	FGFR2-CTNNA3	HIGH EXPRESSION
18	NTRK2-CNTLN	HIGH EXPRESSION
19	MET Exon 14 Skipping	HIGH EXPRESSION
20	MET Exon 14 Skipping	HIGH EXPRESSION
21	EML4-ALK	HIGH EXPRESSION
22	APP-NRG1	HIGH EXPRESSION
23	ADAM9-FGFR1	HIGH EXPRESSION
24	FGFR2-NRBF2	HIGH EXPRESSION
25	LMNA-NTRK1	HIGH EXPRESSION
26	EML4-NTRK3	HIGH EXPRESSION
27	MET Exon 14 Skipping	HIGH EXPRESSION

4. PD-L1 expression in Fusion Positive samples

NTRK fusions	Co-occurring mutations	ALK fusions	Co-occurring mutations	NRG1 fusions	Co-occurring mutations
STOM-NTRK2	TP53	EML4-ALK	TP53	SLC3A2-NRG1	TP53
TPM3-NTRK1	RNF43	EML4-ALK	KMT2D	VDAC1-NRG1	TP53
TPM3-NTRK1	TP53	EML4-ALK	FGFR2	VDAC1-NRG1	TP53
TPM3-NTRK1	FBXW7			APP-NRG1	ARID1A
EML4-NTRK3	RNF43				
EML4-NTRK3	ARID1A				
EML4-NTRK3	TERT				
NTRK2-CNTLN	TP53				
NTRK2-CNTLN	NFE2L2				
NTRK2-CNTLN	PTEN				
LMNA-NTRK1	KMT2D				
LMNA-NTRK1	TP53				
LMNA-NTRK1	TP53				
LMNA-NTRK1	RNF43				
LMNA-NTRK1	FBXW7				
LMNA-NTRK1	POLE				
LMNA-NTRK1	ARID1A				
LMNA-NTRK1	ARID1A				
NTRK2-CDC14B	FBXW7				
NTRK2-CDC14B	TP53				
NTRK2-CDC14B	TP53				

6. Co-occurring driving mutations found on cases with select therapeutically actionable fusions

